

BUFFERS AND REAGENTS

Part A: For S-30 prep

1. Growth media (4L)

- 22.4g KH₂PO₄
- 115.6g K₂HPO₄
- 40g yeast extract

Divide into 3 1L flasks, 5-100ml flasks, and 5-100ml bottles, autoclave. Prior to inoculation, add 40ml of 25% filtered glucose per litre of media.

2. Wash buffer (1L)

- 10mM Tris-acetate, pH 8.0
- 14mM Mg(acetate)2
- 60mM KCl
- 6mM B-mercaptoethanol (just prior to use)
- 50(g/mL PMSF (just prior to use)

3. Lysis buffer (100ml)

- 10mM Tris-acetate, pH 8.0
- 14mM Mg(acetate)2
- 60mM KCl
- 50(g/mL PMSF (just prior to use)

4. Preincubation mix (10ml)

- 3.75ml of 2M Tris-Ac, ph 8.0 (make up fresh)
- 213uL of 1M Mg (acetate)2
- 750uL of 19 aa's - met (1mM each)
- 37.5uL of 20mM methionine
- 600uL of 100mM ATP
- 200mg of phosphoenol pyruvate (PEP)
- 50 units pyruvate kinase
- 75uL of 1M DTT
- H₂O to 10 mL

5. Dialysis buffer (8L)

- 10mM Tris-Acetate, pH 8.0
- 14mM Mg(acetate)2
- 60mM K(acetate)
- 1mM DTT (just prior to use)

Part B: for mfrs prep:

1. TB (2L)
 - 24g bactotryptone
 - 48g yeast extract
 - 8ml glycerol

Autoclave, allow to cool to < 60(C, then add 200ml of sterile, filtered 0.17M KH₂PO₄, 0.72M K₂HPO₄.

2. Buffer A (100ml)
 - 10mM Tris -Acetate, pH 8.0
 - 14mM Mg (acetate)2
 - 60mM K (acetate)
 - 0.1mM DTT (just prior to use)
 - 0.5mM PMSF (just prior to use)
3. Buffer B (200 mL)
 - 10mM Tris-Acetate, pH 8.0
 - 14mM Mg (acetate)2
 - 60mM K (acetate)
 - 0.5% Triton X - 100
 - 0.1mM DTT (just prior to use)
4. Buffer C (100ml)
 - 10mM Tris- Acetate, pH 8.0
 - 14mM Mg (acetate)2
 - 60mM K (acetate)
 - 0.1mM DTT (just prior to use)

Part C: 2.5 X reaction mixture:

FINAL	STOCK	VOLUME/4mL of 2.5X (uL)
35mM Tris-Acetate pH 8.0	2M	175
190mM K(glutamate)	2M	950
30mM NH ₄ OAc	5M	60
2mM DTT	1M	20
12mM Mg(OAc)2	1M	120
40uM 19aa's- met	1M	400

2mM ATP	100mM	200
0.5mM each of CTP, UTP, GTP	15mM	330
20mM phosphoenol pyruvate	400mM	500
0.1mg/mL E.coli tRNA	10mg/mL	100
35mg/mL PEG 8000	400mg/mL	875
20 ug/mL folinic acid	4000ug/mL	50
2mM IPTG	100mM	200
H2O to 4mL		